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(54)【発明の名称】 核酸の単離方法

(57)【要約】

本発明は、サンプルを界面活性剤および固体支持体に接觸させ、これによって上記サンプル中の可溶性核酸がこの支持体に結合し、次いでこの支持体を結合した核酸とともに上記サンプルから分離することからなる、サンプルから核酸を単離する方法を提供する。本発明の方法を、DNAを単離するのに用いる場合、同一のサンプルからRNAを単離する追加工程と都合良く組み合わせることができる。

(2)

RNA

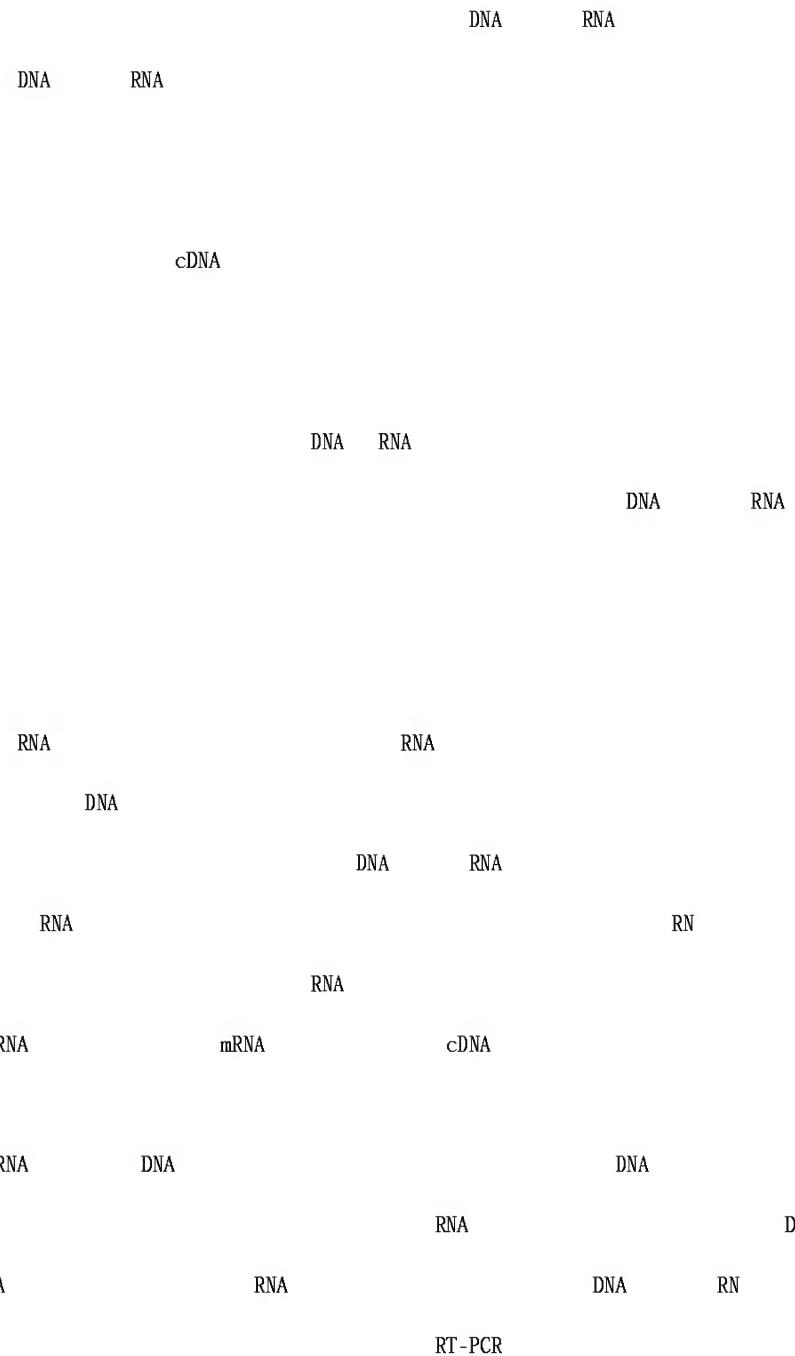
DNA

RNA

0.2 - 30% w/v

(3)

RNA



NA LiC1 DNA R

mRNA	50	300	A
			(polyadenylated)
(A) ⁺ mRNA	RNA	95 %	R
NA	RNA		A

RNA

mRNA

(6)

dT

mRNA

(A)⁺RNA

mRNA

RNA

(A)⁺RNA

microfuge

RNA

LiCl

LiDS/SDS

dT

-K

mRNA

15

mRNA 30

mRNA

mRNA

mRNA

mRNA

GTC

sarkosyl

GTC-

RN

4M GTC

DNA

US-A-5,234,8

(7)

WO 91/12079

RNA

PCR

PCR

RNA

DNA

DNA RNA

DNA DNA

(8)

cDNA

DNA

RNA

RNA

PCR

(9)

(Banerjee S K et al. 1995 Biotechniques 18:769-773)

20

0.5

(SDS)

0.2 30 %

0.5 30 %

0.5 15 %

10 %

1.0 % 0.5 %

(10)

0.1

M 250 500 mM

50 mM

1mEDTA 1mEGTA

10 mM

DTT

-

100mM -HC1 pH 7.5

10mM EDTA

2% SDS

100mM C1 pH 7.5

10mM EDTA

5% SDS

10mM NaC1

100mM C1 pH 7.5

500mM LiC1

10mM EDTA

1% LiDS

DNA-

III
III
10 III
2.8 III 4
.5 III
5%

US-A-4336173

Qiagen Pharmacia

Serotec Dyno Particles AS Lillestrøm Norway

Sintef

EP-A-106873

D

YNABEADS

Dynal AS Oslo Norway

4,336,173 4,459,378

4,654,267

DNA RNA DNA

DNA

10mM -HCl pH 8.0/10mM NaCl

PCR

DNA

DNA

DNA

```

graph LR
    DNA1[DNA] -- "transcription" --> RNA1[RNA]
    RNA1 -- "translation" --> PROTEIN[PROTEIN]
    DNA2[DNA] -- "reverse transcription" --> RNA2[RNA]
    RNA2 -- "replication" --> DNA3[DNA]
    
```

The diagram illustrates the central dogma of molecular biology. At the top, the word "DNA" is written twice. Below the first "DNA", the word "RNA" is written. An arrow labeled "transcription" points from the first "DNA" to the "RNA". Below the "RNA", the word "PROTEIN" is written. An arrow labeled "translation" points from the "RNA" to the "PROTEIN". At the bottom, the word "DNA" is written twice. Below the first "DNA", the word "RNA" is written. An arrow labeled "reverse transcription" points from the first "DNA" to the "RNA". Below the "RNA", the word "replication" is written. An arrow labeled "replication" points from the "RNA" back to the second "DNA".

DNA

```

graph TD
    DNA[DNA] -- "transcription" --> RNA1[RNA]
    RNA1 -- "translation" --> Protein[Protein]
    RNA1 -- "reverse transcription" --> cDNA[cDNA]
    cDNA -- "replication" --> DNA
    RNA2[RNA] -- "replication" --> RNA3[RNA]
    RNA3 -- "reverse transcription" --> cDNA2[cDNA]
    cDNA2 -- "replication" --> DNA
  
```

The diagram illustrates the flow of genetic information. At the top, 'DNA' is shown with an arrow labeled 'transcription' pointing to 'RNA'. This RNA then undergoes 'translation' to produce 'Protein'. Simultaneously, the same 'RNA' undergoes 'reverse transcription' to produce 'cDNA'. 'cDNA' then undergoes 'replication' to produce more 'DNA'. Below this, another 'RNA' molecule undergoes 'replication' to produce 'RNA3'. 'RNA3' then undergoes 'reverse transcription' to produce 'cDNA2'. Finally, 'cDNA2' undergoes 'replication' to produce more 'DNA'.

The diagram illustrates a DNA double helix structure. The outer edges of the helix are labeled "DNA". Along the left side, the sequence "NA LiC1" is written vertically. Along the right side, the sequence "RNA GTC" is written vertically. The interior of the helix is labeled "RNA" in the upper portion and "DNA" in the lower portion.

(15)

RNA
DNA
RNA
RNA DNA

RNA RNA RNA

RNA dT

— DNA
OD nM 0.100
0.427 257.6 nM 0.292 236.4 n

M

— DNA
Hind III
— PCR PCR

Hind III PCR

— PCR Hind III

d III

— Hind III

DNA Dynabeads DNA DIRECT

DNA 10 1

DNA Dynabeads DNA DIRECT

DNA Dynabeads DNA DIRECT

DNA

Hind III

II Dynabeads DNA DIRECT DNA 20 g

DNA 200 ng AMXY PCR

— Dynabeads DNA DIRECT

DNA DIRECT 10 1

DNA 10% DNA

PCR 20% Hind III

III M 100 bp L

—

EDTE

DNA DIRECT

A B 10 L

DNA

EDTA Dynabeads DNA DIRECT

DNA 10% DNA

PCR 20%

— Dynabeads DNA DIRECT

1 100% DNA

DNA DIRECT A B

DNA 10% DNA

PCR 20% II 10⁵

Daudi Dynabeads DNA DIRECT

DNA 120 1 DNA 1

PCR 20%

DNA Hind III PCR

100 bp

10 Dynabeads DNA DIRECT

A DNA DIRECT PCR 20% DNA

M 100 bp B

20ng DNA C

PCR

11 mRNA Dynabeads DNA DIRECT mRNA Dynabe
ads DNA DIRECT DNA Dynabeads oligo(dT)25
100 DNA DIRECT Dynabeads
DNA mg
mg
10mg mRNA
DNA
DNA
RNA

12 (A) (B) (C) (D)
DNA PCR

DNA 200 1 DNA DIRECT 20%
DNA 10% RPC
PCR 2.5% DNA
5% 16S rRNA DNA
DNA 18S rDNA trn
L B15C
DNA
PCR

DNA

4 x 10⁶ HL 60 PBS
10 1 PBS 0.1 ml 5% SDS/10 mM
TrisCl pH 8.0/1 mM EDTA] に再懸濁させたトシリ活性化 Dynabeads® M-
280 DYNAL A/S
一ズすることにより得られる 1 mg の Dynabeads® M-280* を加えた。これに、

1ml
トし、その後 DNA を結合した Dynabeads® を、磁石に引き付け、液相を除

1mM EDTA	50mM NaCl/10mM TrisCl pH 8.0/			
65	0.1 ml			
	DNA			
	DNA	OD ₂₆₀ /OD ₂₈₀	1.72	
TE	DNA	1.7	1.9	DNA
OD ₂₆₀				
50 g/ml	OD ₂₆₀ = 1.0	OD ₂₆₀	0.436	0
.1ml	10 mM	2.18 g	DNA	
DNA		2.67 g	82%	
				>20 kb

表 1

PERKIN-ELMER LAMBDA BIO UV/VIS 分光器
アプリケーション番号 3 : 260 / 280 NM 比

試 料	サイクル	波 長	デーテ	単 位
	15:50	オートゼロ		
004	15:56	260.0 nm	0.436	ABS
		280.0 nm	0.253	ABS
		比	1.723	RAT

DNA

1	EDTA	50	1	5% SDS	1	PBS
の 50 μg の Dynabeads® M-280* を加えた。この溶解物を、1 分室温でイン						
0.5 ml	TrisCl pH 7.5					
DNA						

0.5 ml 10mM TrisCl pH 7.5

DNA 40 1 TE 10mM TrisCl pH 8.0/1 mM EDTA

1 PCR GAPDH PCR

PCR

50 1 PCR 10 1

+++ DNA		
<u>溶解バッファ</u>	<u>洗浄バッファ</u>	<u>結果</u>
2% SDS	50 mM NaCl/1 x TE	+++
2% SDS/1 x TE	50 mM NaCl/1 x TE	+++
2% SDS/1 x TE/10 mM NaCl	50 mM NaCl/1 x TE	+++
5% SDS	50 mM NaCl/1 x TE	+++
5% SDS/1 x TE	50 mM NaCl/1 x TE	+++
5% SDS/1 x TE/10 mM NaCl	50 mM NaCl/1 x TE	+++
1% LiDS/10 x TE/0.5 M LiCl	50 mM NaCl/1 x TE	+++
1% LiDS/10 x TE/0.5 M LiCl	150 mM LiCl/1 x TE	+++
5% LiDS	150 mM LiCl/1 x TE	+++
5% SDS	150 mM LiCl/1 x TE	+++
1% サルコシル	150 mM LiCl/1 x TE	+++

1 x TE 10mM TrisCl pH 8.0/1 mM EDTA 10 X TE 100 mM TrisCl
pH 8.0/10 mM EDTA

実施例1の操作を辿ると、未被膜 Dynabeads® M-450 (Dynal A/S, オスロ、

CD2 DNA

50 1 50 1 PB

S 150mM NaCl/10mM Na₂HPO₄ pH 7.4 10 1 4 x 10⁶
 の Dynabeads® M-450 Pan-T (CD2) (Dynal AS, オスロ、ノルウェーより入手)

30

/

/ 200 1 PBS 200
 g の Dynabeads® M-280* (同上) および 200 μl の溶解バッファ [100mM

Tris-HCl pH 8.0/500mM LiCl/10mM EDTA pH 8.0/1% LiDS

DNA/

DNA/ 200 1 [10mM Tris-HCl
 pH 8.0/150mM LiCl/1mM EDTA pH 8.0] 50 1

65

1 GAPDH PCR

DNA

DNA m1 EDTA Dynabeads DNA
 DIRECT Dynal A/S,

Dynabeads® M-280* と等価のビーズを含有したキット) を用いて、同じ血液

10 1 DNA 65

DNA Dynabeads

Dynabeads DNA DIRECT DNA

DNA 0.2%

DNA 10 1 (5m1 0

.2%)

DNA John John S.W.M. G Weitzner R Rose

n C.R.scriver 1991 A Rapid Procedure for Extracting Genomic DNA

from Leukocytes Nuc1 Acid Res 19(2): 408

Dynabeads DNA DIRECT 200 1 Dynabeads DN

A DIRECT 10 1 1.5 ml

(/ 200 g Dynabeads)

DNA Dynabeads

DNA/Dynabeads Dynal E(Magnetic Particle

Collector E) (MPC-E)

Dynal MPC

10 1

TE pH 8.0

65

DNA

DNA 1.5%

1 x TAE DS34

667

I 1

()

DNA ()

Hind III 23.13 kb

DNA 20 kb

DNA DIRECT

ACD DNA 10%

DNA 200 ng X-Y

(X-Y homologous amelogenin) (AMXY) (Akane A. K. M

atsubara H Nakamura S Takahashi K Kimura 1994 Purification

of Highly Degraded DNA by Gel Filtration for PCR BioTechniques 16(2):

235-238) amplicon PCR

DNA DIRECT

PCR 50 1 10 x PCR Perkin Elmer

1 x dNTP Pharmacia 0.2mM

(ampliataq) (Perkin

Elmer pmol AMXY-1F 5'-CTGA

TGGTTGGCCTCAAGCCT-GTG-3' AMXY-4R 5' -TTCATTGTAAGACCAAAGCAAACA-3'

PCR Perkin Elmer GeneAmp PCR System 9600

AMXY PCR 94 38 x[94 30

55 30 72] 72 10

1.5%

DS34 667

J X-Y

DNA (Akane et al 1994)

) 908 bp X 719 bp Y

II X Y

Dynabeads DNA DIRECT

DNA

DIRECT DNA

DNA PCR DNA DIRECT

DNA PCR

溶解／結合 バッファ：

0.5 M LiCl
1 % LiDS
0.1 M TrisCl pH 7.5
10 mM EDTA
5 mM ジチオトレイクトール (DTT)
0.15 M LiCl
10 mM Tris-HCl pH 8.0
1 mM EDTA

洗浄バッファ：

DNA

Dynal AS

Dynabeads DNA DIRECT

DNA

A: 3.6×10^6 /ml B: 2.6×10^6 /ml

200 1 Dynabeads DNA DIRECT

1.5 ml

DNA Dynabeads

DNA/Dynabeads

Dynal

E(MPC-E)

Dynal MPC

40 1 TE pH 8.0

PCR

10%

DAPDH

amplicon

PCR

PCR

50

10 x PCR

(Perkin E

Elmer) 1 x

dNTP Pharmacia 0.2mM

(ampliTaq) (Perkin Elmer)

(25)

	pmol	GAPDH-Forward (5'-ACAGTCCATGCCATCAC				
TGCC-3')		GAPDH-Reverse (5'-GCCTGCTTCACCACCTCTTG-3')				
PCR	Perkin Elmer GeneAmp PCR System 9600					GAPDH
	PCR	94	34 x[94	30	61	30
72]	72	10			
DNA	PCR					1.5%
		50	1	10	1	D
NA	50%			1 x TAE		
	DS34			667		

DNA

Dynabeads DNA DIRECT Dynal AS

EDTA

DNA

DNA 200 1 Dynabeads DNA DIRECT

1 10 1

1.5 ml

DNA Dynabeads

DNA/Dynabeads Dynal E (MPC-E)

Dynal MPC

20 40 1 TE pH 8.0

40

1

20 1

10% (20%)

(DAPDH) amplicon PCR

PCR Dynabeads TE

PCR 50 1 10 x PCR

Perkin Elmer 1 x dNTP

(Pharmacia) 0.2 mM (

ampliTaq (Perkin Elmer) pmol G

APDH-Forward (5' -ACAGTCCATGCCATCACTGCC-3') GAPDH-Reverse (5' -GCCTGCT

TCACCACCTCTTG-3') PCR Perkin Elmer GeneAmp PCR S

ystem 9600 GAPDH PCR 94

34 x[94 30 61 30 72] 72 10

50 1 10 1 DNA 25%

50% DNA PCR

1.5%

1 x TAE DS34

667

1 PCR

DNA 20% 10 1

10%

. M Georgesz A.M Lew 1993 FoLT PCR: A Simple PCR Protocol for Amplifying DNA Directly from Whole Blood BioTechniques 14(3): 238-243)

DNA DIRECT ACD(II) CPD

_____ DNA _____

Dynabeads DNA DIRECT Dynal AS

EDTA DNA

20 +4

DNA

200 1 Dynabeads DNA DIRECT

1.5 ml

DNA Dynabeads

DNA/Dynabeads Dynal E (MPC-E)

Dynal MPC

40 1 TE pH 8.0 PCR

Dynabeads TE

10%

DAPDH amplicon PCR
DNA PCR

1.5% 50 1 10 1
 DNA 50% 1 x TA
 E DS34 667
 I
 +4 -20
 Dynabeads DNA DIRECT
 DNA
 10 1
 1.5 ml 40 1 PBS
 90 DNA Dynabeads DNA DIRECT
 10%
 DAPDH amplicon PCR
 DNA PCR 1.5
 % 50 1 10 1
 DNA 50% 1 x TAE
 DS34 667
 II
 DNA PCR

DNA

1

DNA DIRECT DNA
 200 1 Dynabeads DNA
 DIRECT 1.5ml

DNA Dynabeads
 DNA/Dynabeads Dynal 1 E (MPC-E)

Dyna1 MPC

40 1 TE pH 8.0

Dynabeads PCR

10% DAPDH

amplicon PCR

DNA PCR 1.5%

50 1 10 1 D

NA 50% 1 x TAE

DS34 667

I

()

I DNA

1 DNA DIRECT

1 DNA

1 PCR

1 DNA

4 x 10⁵ Daudi cells DNA DIRECT

DNA 4 x 10⁵

DNA Dynabeads DNA DIRECT ml

ml

DNA/Dynabeads 120 1 TE

120 1 1 D

(30)

APDH amplicon PCR
DNA PCR 1.5%
50 1 10 1 D

NA 10% 1 x TAE
DS34 667

II 120

PCR

Dynabeads DNA DIRECT Dynal AS

DNA

200 1 Dynabeads DNA DIRECT 1.5 ml

ds

DNA Dynabeads DNA Dynabea

DNA/Dynabeads Dynal E(MPC-E)

Dynal MPC

10

1 Dynabeads

DAPDH amplicon PCR
PCR 1.5%
50 1 10 1

1 x TAE

DS34

667

10 PCR

mRNA	DNA DIRECT	DNA	
mRNA	100	0.75m1	
/	DNA DIRECT Dynabeads	DNA-Dynabeads	Dyna1
MPC-E		10mg	
DNA DIRECT	DNA	(11)	
mg	Dynabeads	(dT)25	Dy
nabeads		mRNA	
	mRNA-Dynabead	MPC-E	
		Li	
DS	0.75m1		
	mRNA-Dynabead		
LiDS			
	mRNA	20 1 mM Tr i	
s-HCl pH 7.5	65	Dyn	
abeads		1.0%	
		11	
EtBr-	DNA	rRNA	
RNA			
11	10	DNA	mRNA

DNA	DYNAL	mRNA	DNA DIRECT
DNA			
溶解／結合 バッファ:		0.5 M LiCl	
		1 % LiDS	
		0.1 M TrisCl pH 7.5	
		10 mM EDTA	
		5 mM ジチオトレイトール (DTT)	
L i D S 含有洗浄バッファ:		0.15 M LiCl	
		0.1 % LiDS	
		10 mM Tris-HCl pH 8.0	
		1 mM EDTA	
洗浄バッファ:		0.15 M LiCl	
		10 mM Tris-HCl pH 8.0	
		1 mM EDTA	

<u>DNA</u>			
<u>PCR</u>		<u>DNA</u>	
(E.Coli)		<u>Bacillus cereus</u>	LB
37		(Agrobacterium	
<u>tumefaciens</u>) YEB		40 28 (Sambrook J et al	
. 1989 Molecular Cloning: A Laboratory Manual 2nd ed. Cold Spring Ha			
rbour Laboratory NY.)		(Prochlo	
rthrix)	NIVA	14 18 20	
Norwegian Institute of Water Research 1991			
20	450,000	DNA	

mg 20mg

DNA

Saccharomyces cerevisiae

IMR (Epp)

Key R et al. 1967 Exp Mar Biol Ecol 1 191-208)

Arabidopsis thaliana (Hordeum vulgare)

Perca fluviatilis

mg 30 100mg 100

400 mg DNA DNA

foreceps

(Kontes Scientific

Instruments Vineland New Jersey USA)

DNA

Dynabeads DNA DIRECT Dynal AS

DNA 200 1 D

Dynabeads DNA DIRECT / 200 g Dynabeads

1.5 ml 15 DNA Dynabeads

65 15

DNA/Dynabeads Dynal E (MPC-E)

Dynal MPC

40 1 TE pH 8.0

	65	
	DNA	
DNA		1.5%
1 x TAE		DS34
	667	
溶解／結合バッファー：		
0.5 M LiCl		
1% LiDS		
0.1 M TrisCl pH 7.5		
10 mM EDTA		
5 mM ジチオトレイトール (DTT)		

洗浄バッファー：	
0.15 M LiCl	
10 mM Tris-HCl pH 8.0	
1 mM EDTA	

DNA DIRECT	
/	-
	DNA

Sambrook J et al. 1989

A-5 Sigma Chemicals Co.

St Louis USA

DNA Scot O.R. Bendich A.J. 1994 "Plant Molecular Biology Manual" page D1: 1-8 Kluwer Academic Publisher Belgium

PCR

PCR	DNA	DNA
R		PC
15 pmol	200 M dNTP 10mM Tris-HCl pH 8.8 1	
.5 mM MgCl ₂ 50 mM KC1 0.1% Triton X-100		DynaZyme
	Finnzymes Oy Finland	0.1-5 1 DN
A 50 1	PCR	Perkin Elmer GeneAmp PCR Sy

stem 9600

amplicon

PCR	94-97	DNA	72
-----	-------	-----	----

amplicon

			IUD
Brosius J. et al. 1978 Proc Natl Acad Sci. USA 57 4801			
-4805	<u>E Coli</u>	334 939	16S rRNA

プライマー: CC 5'-TGTAAAACGACGCCAGTCCAGACTCCTACGGGAGGCAGC-3'
CD 5'-CTTGTGCGGGCCCCGTCAATTC-3'

CC	-21 M13		5
		DNA	: 96 1
5	70	30	

18S rRNA			Medlin et al. 1990 Gene 71 491-499
A	B		
94	30	50	72 35

18S rRNA	600 bp.	White et al. 1990 Innis M.A. et al.	
"PCR Products a Guide to Methods and Applications" page 315-32			

2 Academic Press New York			NS3 NS4
---------------------------	--	--	---------

94	30	53	30	72
94	30	50	30	15
72		25		

tRNA	I	Fangan et al. 1994 BioTechn			
iques 16 484-494	C	D			
94	30	55	30	72	30

Arabidopsis thaliana B15C 800 bp

5' -CGGGATCCCTAGGAGACACGGTGCCG-3' ↗ ↘
 5' -GGAATTGGATCGGCGGTCTGAAAC-3' .

94	30	59	30	72	35
----	----	----	----	----	----

Barley B15C 800 bp

5' -CGGATCCCGTCATCCTCTTCTCGCACCCC-3' ↗ ↘
 5' -GGAATTCCCTCTTGGAGGGCAGGTCGGCG-3' .

94	30	60	30	72	35
----	----	----	----	----	----

(Perch)

D-	800-900 bp	Hoelzel et al. 1
991 Mol Biol Evol. 8 475-493	HV2	

5' -GGTGACTTGCAATGTGTAAGTTCA-3' .

96	52	72	30
----	----	----	----

1.5%

1 x TAE DS34

100-1000 ng

DNA (12A)

65 15

DNA 500 ng mg

0.25% DNA

DNA 100-200 ng

300-500ng (12B)

DNA 5

% DNA PCR (12B)

0.5-5% DNA PCR

DNA DIRECT

200-400 ng DNA (12C)

PCR DNA PCR

5% DNA

DNA

65 15

(12D) PCR DNA DNA

PCR 5% DNA

300-500 ng DNA

() DNA 5

% DNA

PCR

Hultman et

al. 1989 Nucleic Acids Res. 17 4937-4946

表 2: 異なる器官からのDNA単離およびPCR増幅

種 ^a	試料 ^b	DNA 収量 ^c	ゲノム ^d Gen.	PCR 生成物 ^d Org.
<u>細菌</u>				
<i>Bacillus cereus</i> ATCC 75	新鮮	+++	16S	+++
<i>E. coli</i> NovaBlue ¹	新鮮	+++	16S	+++
<i>A. tumefaciens</i> GV 310 ²	新鮮	+++	16S	+++
<i>Plantarothrix agardhii</i> N-C 29	新鮮	+++	16S	+++
<i>P. prolificans</i> N-C 320	新鮮	+++	16S	+++
<i>Microsphaeris aruginosa</i> N-C 43	新鮮	+++	16S	+++
<i>M. aruginosa</i> N-C 228/1	新鮮	+++	16S	+++
<i>Anabaena bory</i> N-C 246	凍結	+++	16S	++
<i>Phormidium</i> sp N-C 177	凍結	+++	16S	++
<i>Aphanizomenon</i> sp N-C 103	凍結	+++	16S	+++
<i>P. hollandica</i> N-S/89	凍結	++	16S	+++
<u>菌類</u>				
<i>Corticarius sanguineus</i>	d.fruitb.	+++	16S	+++
<i>Corticarius gentilis</i>	d.fruitb.	++	16S	+++
<i>Russula lutea</i>	d.fruitb.	++	16S	+++
<i>Laccaria bicolor</i>	f.mycel	+	16S	+++
<i>Trichia ochroleuca</i>	f.mycel	++	16S	+++
<i>Verjakinia cultica</i>	f.mycel	++	16S	+++
<i>Periza vesiculosus</i>	f.mycel	++	16S	+++
<i>Saccharomyces cerevisiae</i>	新鮮	+	16S	+++
<u>藻類</u>				
<i>Gyrodinium aureolum</i>	新鮮	+++	16S/16S	+++/+++
<i>Heterocapsa triplex</i>	新鮮	+++	16S/16S	+++/+++
<i>Scrippiella trochidea</i>	新鮮	+++	16S/16S	+++/+++
<i>Ceratium strictum</i>	新鮮	+++	16S	+++
<i>Chlorella vulgaris</i>	新鮮	+++	16S	+++
<i>Clamydomonas reinhardtii</i>	新鮮	+++	16S	+++
<i>Calicium usidaria</i>	新鮮	+++	16S	+++
<i>Chrysotrichum polylepis</i>	新鮮	+++	16S	+++
<u>植物</u>				
<i>Hordeum vulgare</i> (barley)	草子葉類	+++	B1SC/αmL	+++/+++
<i>Arabidopsis thaliana</i>	双子葉類	+++	B1SC/αmL	+++/+++
<u>脊椎動物</u>				
<i>Perca fluvarialis</i> (perch)	魚類	cp.	D-ループ	+++

^a *A. tumefaciens* = *Agrobacterium tumefaciens*.^b *P. hollandica* = *Prochlorothrix hollandica*.^c d.fruitb. = 乾燥子実体, f.mycelia = 新鮮な菌子体, cp. = ひれ.^c 標準的なフェノール/クロロホルム単離に関する大体のDNA収量

+++; > 80 %, ++; > 10 %, +; > 1 %, nL = 試験せず

^d Gen. = ゲノムDNA, Org. = 葉緑体からのオルガネラDNA(藻類および植物)およびミトコンドリア(魚類)^e 実施例3に記載したアンブリコン

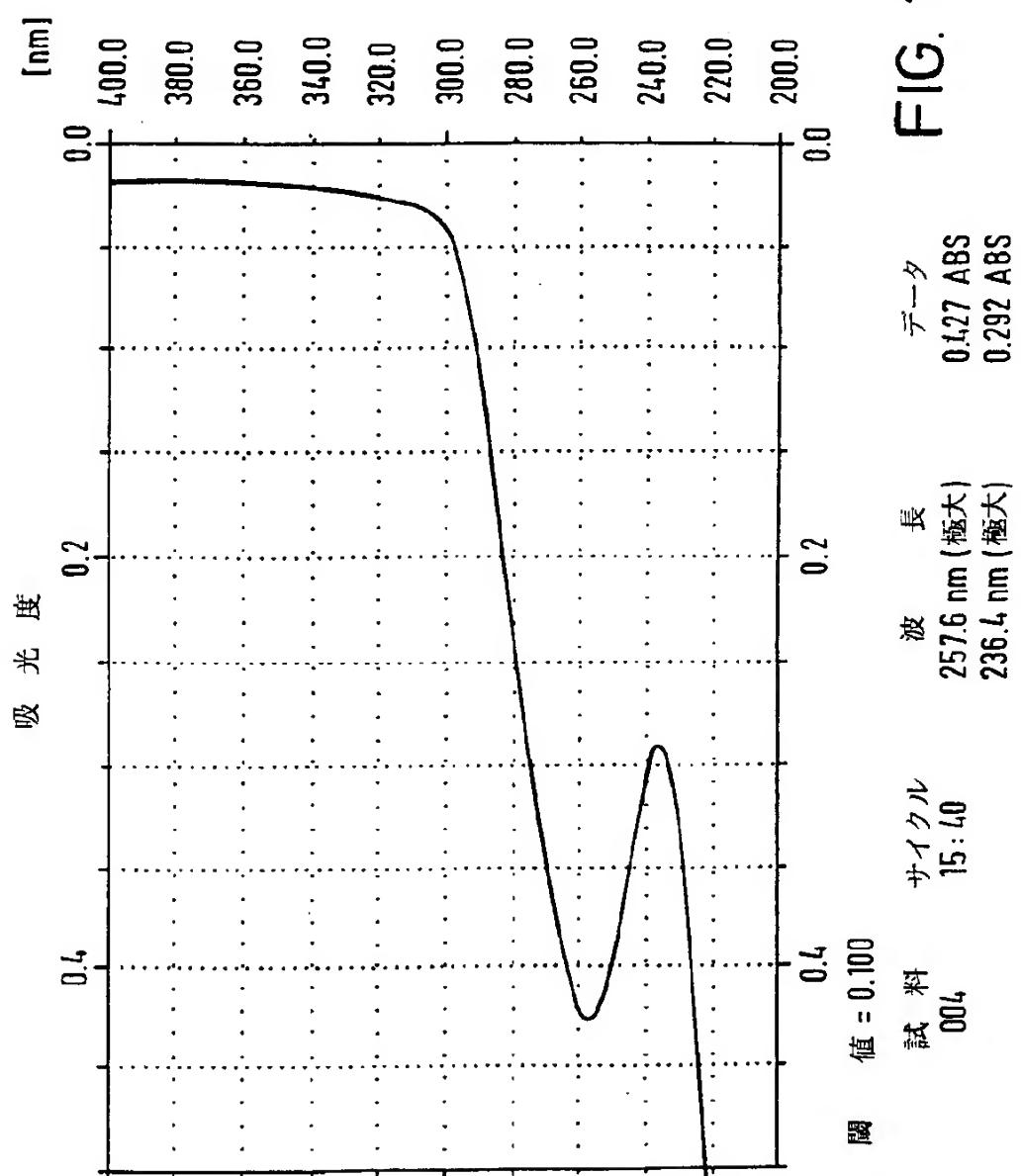
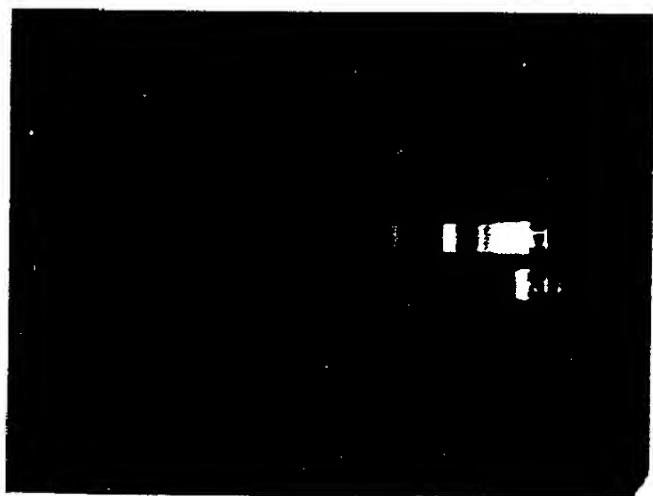


FIG. 1



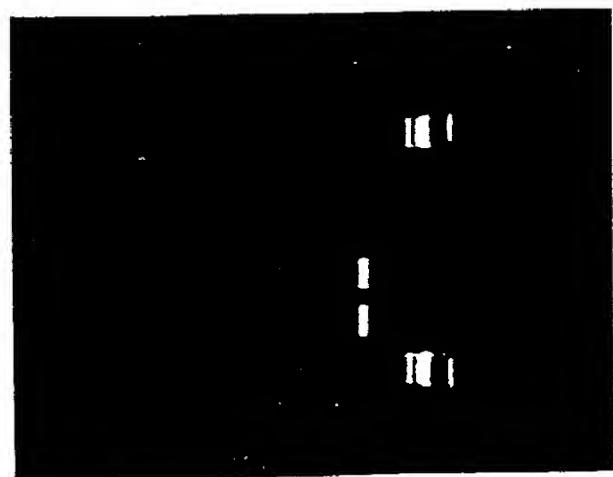
1 2

FIG. 2



1 2 3

FIG. 3



1 2 3 4 5 6

FIG. 4

パネル I
パネル II

1 2 3 4 5 6 7 . 1 2 3 4 5

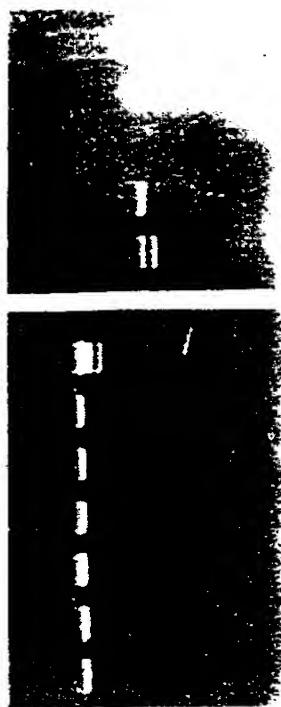


FIG. 5

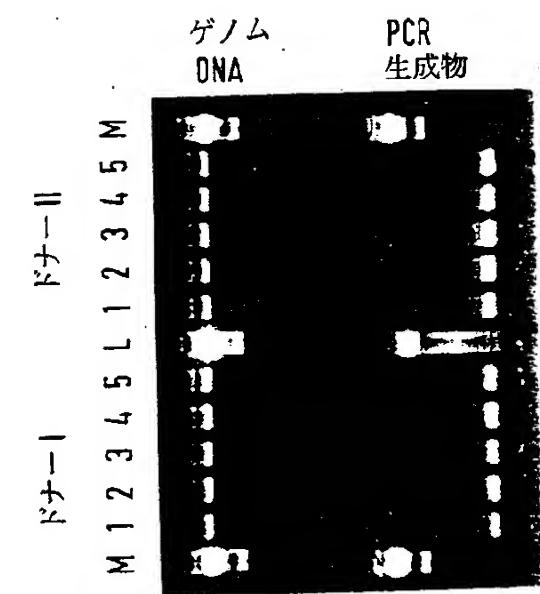


FIG. 6

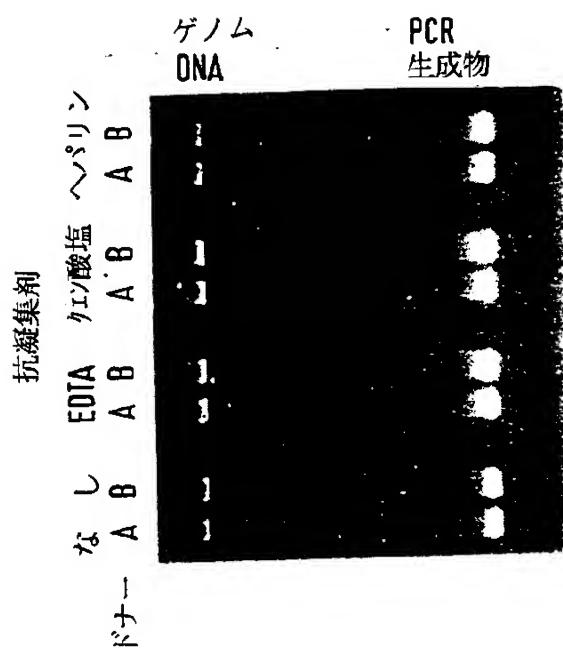


FIG. 7

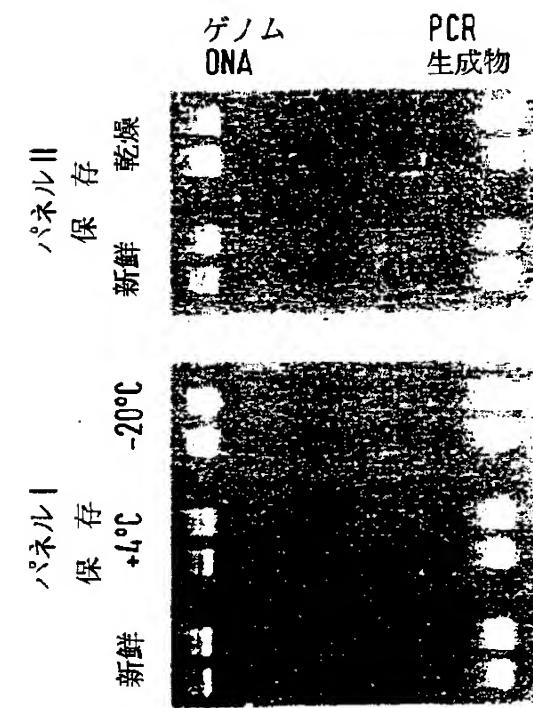


FIG. 8

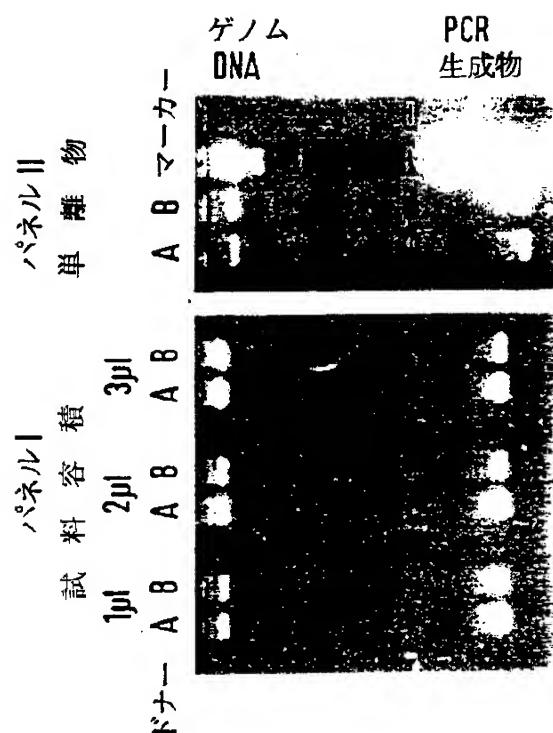


FIG. 9

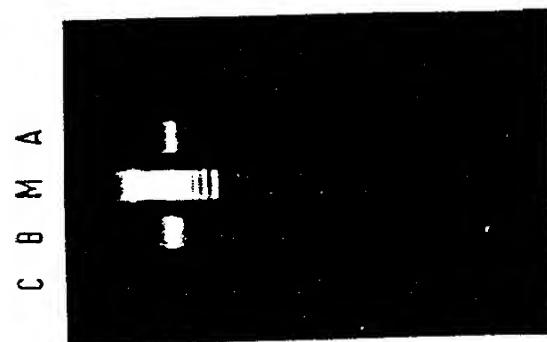


FIG. 10

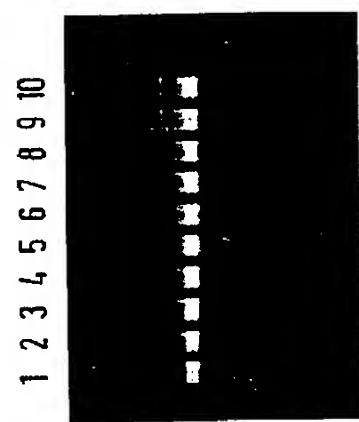


FIG. 11

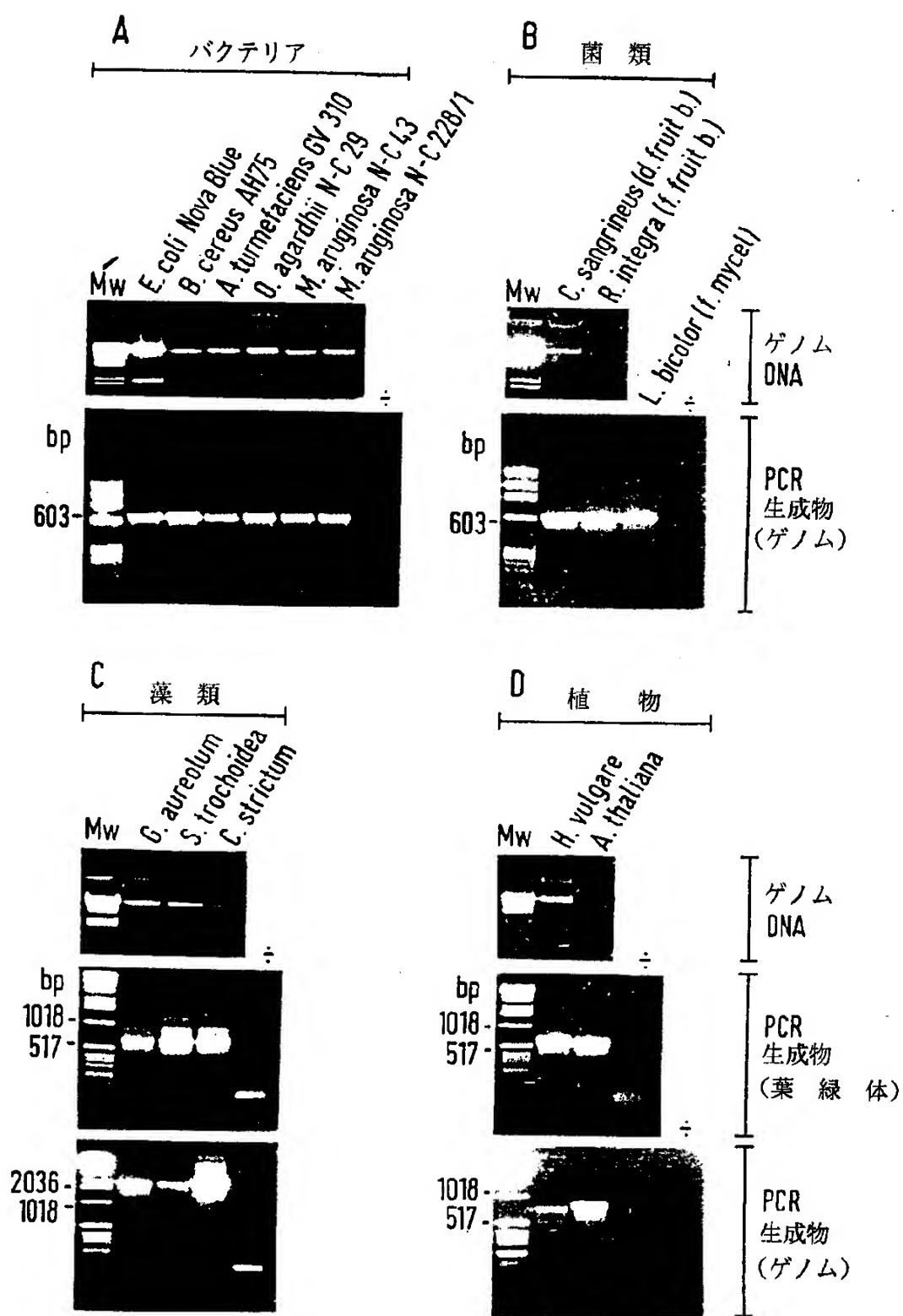


FIG. 12

INTERNATIONAL SEARCH REPORT

International Application No
PCT/GB 95/02893

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/10

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO,A,93 25912 (MEDICAL RESEARCH COUNCIL) 23 December 1993 *see the whole patent* ---	1-18
X	JOURNAL OF APPLIED BACTERIOLOGY, vol. 74, 1993, pages 78-85, XP002007385 K. SMALLA ET AL.: "Rapid DNA extraction protocol from soil for polymerase chain reaction mediated amplification" *see the whole article* ---	1-18 -/-

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
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- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
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- "&" document member of the same patent family

Date of the actual completion of the international search

3 July 1996

Date of mailing of the international search report

02.08.96

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Marie, A

INTERNATIONAL SEARCH REPORT

International Application No
PCT/GB 95/02893

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	APPLIED AND ENVIRONMENTAL MICROBIOLOGY, vol. 60, no. 5, 1994, pages 1572-1580, XP002007386 N.I. MORE ET AL.: "Quantitative cell lysis of indogenous microorganisms and rapid extraction of microbial DNA from sediment" *see the whole article* ---	1-18
X	EMBO JOURNAL, vol. 4, no. 4, 1985, pages 913-918, XP002007387 D.A. JACKSON ET AL.: "A general method for preparing chromatin containing intact DNA" *see the whole article* ---	1-18
X	EMBO JOURNAL, vol. 3, no. 8, 1984, pages 1837-1842, XP002007388 P.R. COOK : "A general method for preparing intact nuclear DNA" *see the whole article* ---	1-18
X	EXPERIMENTAL CELL RESEARCH, vol. 190, 1990, pages 294-295, XP002007389 G. KONAT ET AL.: "Rapid isolation of genomic DNA from animal Tissues" *see the whole article* ---	1-18
X	ANALYTICAL BIOCHEMISTRY, vol. 164, 1987, pages 207-213, XP002007390 P.M. GLEE ET AL.: "Methods for DNA extraction from <i>Candida albicans</i> " *see the whole article* -----	1-8

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No
PCT/GB 95/02893

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO-A-9325912	23-12-93	AU-B-	4343993	04-01-94
		AU-B-	4344093	04-01-94
		WO-A-	9325709	23-12-93

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